

**TITLE: The Mycorrhizal Tragedy of the Commons**

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N.H. performed the experiment, analyzed data, and drafted the manuscript. O.F. developed the model based on experimental data. L.T., J.M., T.N., J.F., and L.E. contributed to experimental design and interpretation of data. All authors contributed to the manuscript text.

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The data supporting the results and the model script will be archived in an appropriate public repository, and the data DOI will be included in the article.

## 38 **ABSTRACT**

39 Trees receive growth-limiting nitrogen in exchange for allocating carbon to mycorrhizal  
40 symbionts, but supplying the fungi with carbon can also cause nitrogen immobilization,  
41 which hampers tree growth. We present results from field and greenhouse experiments  
42 combined with mathematical modelling, showing that these are not conflicting outcomes.  
43 Mycorrhizal networks connect multiple trees, and we modulated C provision by strangling  
44 subsets of trees, assuming that carbon supply to fungi was reduced proportionally to the  
45 strangled fraction. We conclude that trees gain additional nitrogen at the expense of their  
46 neighbors by supplying more carbon to the fungi. But this additional carbon supply  
47 aggravates nitrogen limitation via immobilization in the shared fungal biomass. We illustrate  
48 the evolutionary underpinnings of this situation by drawing on the analogous *tragedy of the*  
49 *commons*, where the shared mycorrhizal network is the commons, and explain how rising  
50 atmospheric CO<sub>2</sub> may lead to greater nitrogen immobilization in the future.

## INTRODUCTION

In boreal forests, ectomycorrhizal fungi (EMF) contribute significantly to tree nitrogen (N) acquisition, which is frequently the growth limiting factor in this biome (Högberg *et al.* 2017). But mycorrhizal N is acquired at the cost of photosynthetic carbon (C) (Colpaert *et al.* 1996; Corrêa *et al.* 2008). This is the basis for mycorrhizal trade, but despite the fact that it is one of the most widespread and influential symbioses in boreal forest ecosystems, the ecological nature of this exchange has not been settled (Johnson *et al.* 1997; Alberton *et al.* 2005; Franklin *et al.* 2014; Terrer *et al.* 2019). This represents a critical weak point in any predictions of ecosystem responses to future perturbations of C or N dynamics (Alberton *et al.* 2007; Högberg *et al.* 2017).

Ectomycorrhizal symbioses can vary from mutualism to competition to parasitism, depending on prevailing growth conditions (Johnson *et al.* 1997; Alberton *et al.* 2007; Ågren *et al.* 2019). Under N limitation, host plants have been observed to continue supplying their ectomycorrhizal partner with C, and even increasing the C investment, despite diminishing N returns (Corrêa *et al.* 2008, 2010). If N availability is amended via fertilization, however, EMF transfer a greater proportion of their absorbed N (Näsholm *et al.* 2013). N is thus withheld under conditions of limiting availability, and the host tree cannot unlock it by supplying the EMF with more C, because such an investment results in further diminishing N returns. The eco-evolutionary explanation is that each fungal individual competes with other EMF symbionts of the same plant and can gain a larger share of the plant's C supply by increasing its N export, until its own remaining N matches its C supply (Näsholm *et al.* 2013). Conversely, a larger C flux from the plant allows the fungus to use a greater proportion of the N it absorbs from the soil, as dictated by the stoichiometric requirements of fungal biomass and growth (Alberton *et al.* 2007; Näsholm *et al.* 2013; Franklin *et al.* 2014). Enhanced EMF growth may initially increase N uptake and, by extension, export to host plants but N availability eventually becomes limiting, whereas N immobilization in fungal biomass continues, leading to a negative feedback on the plant's N uptake (Corrêa *et al.* 2010; Näsholm *et al.* 2013). This sequence has been suggested as a mechanism for observed progressive N limitation in forests under increased atmospheric CO<sub>2</sub> concentrations (Alberton *et al.* 2007; Högberg *et al.* 2017). The presence of such an N-immobilizing feedback

loop raises the question of how the symbiosis can remain stable over evolutionary time scales and how it has survived natural selection.

Here we present an ecological framework to reconcile the observed plant and fungal behaviors summarized above, by recognizing the dual scale of the ectomycorrhizal symbiosis. The critical point is that multiple fungi can colonize the roots of a given tree and that several trees can be connected to the same fungal individual, creating common mycorrhizal networks (Southworth *et al.* 2005). Trees have evolved to maximize their own competitive benefit from the symbiosis at the individual tree scale, but this maximization also has consequences for other trees with whom they share EMF partners at the *network scale*.

The classic paper titled *The Tragedy of the Commons* (Hardin 1968) presents a theory of over-exploitation of shared resources which effectively illustrates the evolutionary underpinnings which have led to the widespread success of a symbiosis in which one partner is in fact maintaining its own resource limitation: Trees do not coordinate their carbon investments within the shared fungal network, but act to increase individual advantages over their neighbors. In Hardin's example, a common pasture was depleted by several herdsmen who all increased the number of cattle they kept there. From each herdsman's perspective, this is the rational course of action, because the cost of the degraded pasture is divided among all users, whereas the individual herdsman receives the entire reward of an extra head of cattle. Analogously, all host trees are competing for enhanced shares of mycorrhizal nutrients, but their combined efforts serve to aggravate the overall nutrient immobilization in fungal biomass.

## **HYPOTHESIS**

We hypothesize that common EMF symbionts reward the host plants that supply the most C by allocating a greater proportion of their total exported N to them. Conversely, the hosts, which share multiple EMF symbionts, reward the partners that supply the most N by releasing a greater proportion of their total C export to those networks. The "tragedy" from the plant viewpoint arises when the total C export to all fungi is so high that it leads to N

immobilization, in which case the proportions cease to matter and all plants suffer. The “tragedy” from the fungal viewpoint arises when exporting more N would reduce their own growth, but exporting less N would reduce their competitiveness for plant C (Näsholm 2013, Franklin 2014).

This hypothesis has specific and predictable implications for plant N uptake in response to C export (mathematically formulated in the Materials & Methods section of this article): A positive linear relationship at the individual plant level and a saturating or hump-shaped relationship at the community level (fig. 1, alternative 3). As stated, the hypothesis should be rejected if a similar relationship between N uptake and C export were observed at both individual and community levels (fig. 1, alternatives 1, 2). Such a result would imply a lack of inter-plant competition for N through a common network.

We conducted two experiments, (in the greenhouse and field conditions), where belowground C flux was reduced by shading and/or stem strangling. Strangling is a treatment whereby belowground C flux is physically restricted by blocking phloem transport. It has been proven to consistently control plant C export to roots and EMF (Björkman 1944; Henriksson *et al.* 2015). Strangling a subset of seedlings growing in the same pot accomplishes two things: 1) the total belowground C flux is decreased, and 2) each seedling’s relative contribution to that flux is altered. Shade treatments were also applied to uniformly reduce total C availability to belowground structures in a subset of pots.

## MATERIALS AND METHODS

### SEEDLING EXPERIMENT

#### Preparation of mycorrhizal inoculum

A culture of *Suillus variegatus* was prepared based on the protocol described in (Vuorinen *et al.* 2015) with a few modifications. Briefly, ½ MMN medium, was prepared. The media contained 1.25 g/L glucose, 5 g/L malt, 0.5 g/L (NH<sub>4</sub>)<sub>2</sub>HPO<sub>4</sub>, 0.5 g/L KH<sub>2</sub>PO<sub>4</sub>, 0.15 g/L MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.05 g/L CaCl<sub>2</sub>·2H<sub>2</sub>O, 0.025 g/L NaCl, 0.02 g/L Fe-EDTA, 0.02 g/L Thiamine-HCl

and the pH value adjusted to 5.8 with NaOH-HCl. As inoculum, plugs (5x5 mm) from the actively growing peripheral zone of *S. variegatus* mycelia, growing on solid ½ MMN agar plates, was used. The mycelia were first cultured in 250 ml ½ MMN medium in 1 L Erlenmeyer flasks, sealed with cotton and aluminum foil for 16 days in a dark incubator at 23°C and rotation speed at 100 r/m. Following this, the liquid culture was homogenized and mixed with silica powder (Sipernat 22S, Algol Chemicals AB) moistened with ½ MMN medium to a moisture content of 70%, by weight (250 g silica in each container). The containers were placed in a dark and ventilated space and the mycelium was allowed to grow for another 28 days.

### **Seedling growth conditions**

On May 10<sup>th</sup>, 2018 (DOY 130), two-year-old *Pinus sylvestris* seedlings were bare-rooted, weighed and potted in a soil mix containing 10 % *S. variegatus* inoculum and 90 % soil (50-50 mixture of peat and commercial non-fertilized potting soil). No measures were taken to exclude fungal species already present on the roots of the seedlings. Each pot (23 cm x 17 cm x 6 cm) contained six seedlings, planted as shown in fig. 1b. Seedling fresh weight was measured before planting. Seedlings were randomly selected to pots and there was no significant difference in initial fresh weight among seedlings that would subsequently be allotted to different treatments ( $P = 0.74$ ).

To allow the seedlings to establish themselves in their pots, they were kept in controlled greenhouse conditions for 53 days. They were then transferred outdoors, first to partial shade for 10 days, to avoid damage to the needles from sudden exposure to direct sunlight, and then into the open. After 18 days in direct sunlight, the experimental treatments were initiated. Thus, the experimental treatments were begun on July 30<sup>th</sup> 2018 (DOY 211), 81 days after the seedlings were potted. Finally, all the pots were transferred back inside the greenhouse for the final month of the study. This was done to avoid loss of <sup>15</sup>N tracer in autumn rains. All pots were watered daily throughout the experiment duration.

### **Shading and strangling treatments**

Half of the pots were covered by individual shade tents constructed of DeWitt UV PE knitted shade cloth (Agriculture Solutions, LLC, Strong, Maine), reducing incoming photosynthetic radiation (PAR) by  $78.8 \pm 4.8$  % (mean  $\pm$  SD).

Within each pot, the six seedlings could be either strangled or left unstrangled, and the treatments were designed so that 0, 1, 5, or 6 seedlings were strangled. Thus, there were four levels of the strangling treatment, and two levels of the light treatment (Fig. 1). This resulted in a total of eight factorial combinations of treatments that were replicated 5 times in a blocked design.

Seedlings were strangled by tightly wrapping iron wire (0.7 mm diameter) around the stems below the lowest branch (Björkman 1944). This method, and modified versions for large trees, have been shown to effectively reduce belowground C flux in *P. sylvestris* and *Pinus ponderosa* (Björkman 1944; Henriksson *et al.* 2015). In Björkman (1944) strangling of 3-year-old pine seedlings for one entire season was shown to strongly reduce root soluble carbohydrate levels as well as ectomycorrhizal colonization rate ( $< 5$  % of root tips, compared to c. 65 % in control seedlings). In that publication, the strangling wire was removed from a subset of the seedlings after three months, resulting in intermediary levels of both measurements.

### **<sup>15</sup>N application**

Thirty-two days after the initiation of shade and strangling treatments, <sup>15</sup>N was applied to the soil surface of each pot (DAY 243), in the form of KNO<sub>3</sub> (Larodan AB, Karolinska Institutet Science Park, Stockholm, Sweden). The total added quantity corresponded to 0.3 g <sup>15</sup>N / m<sup>2</sup>, which was applied in three doses over the course of six days to avoid flushing the system with nitrogen. Each pot thus received a total of 0.012 g <sup>15</sup>N.

To avoid loss of <sup>15</sup>N from the bottom of the pots, as well as isotopic contamination between pots, the pots were placed in individual trays before isotopic label was applied, and remained in these for the duration of the study. Any water that drained out the bottom was used to re-water the same pot using a syringe.

### **Final harvest and sampling**

Three weeks after the <sup>15</sup>N labelling (on DOY 269-276), the seedlings were harvested. All 240 seedlings (6 seedlings x 40 pots) were washed and their roots separated. The needles, roots,

stem, and buds of each seedling were separated, and weighed. For strangled seedlings, the wire was removed before weighing the stem. The material was then dried at 60°C for 48 hours before being weighed again.

The dried needles, roots and stem of each seedling were milled in a chamber mill (IKA-Werke GmbH & Co.KG). Using isotope ratio mass spectrometry, the total C and N contents (% C and % N) and the isotopic enrichment of <sup>15</sup>N was analyzed for each plant compartment.

The current setup allows quantification of the <sup>15</sup>N abundance in the entire seedling biomass, rather than relying on foliage concentration, which is commonly used as a measure of N uptake in field conditions (Hasselquist *et al.* 2016). By measuring each seedling compartment separately (needles, roots, stem), we avoid problems in distinguishing actual N mobilization from a potential shade avoidance response in the trees (Henry & Aarssen 1997), which could shift internal N partitioning toward the foliage.

## Statistical analyses

Seedling dry weights, elemental and isotopic composition, and photosynthetic rates were compared using a standard least squares means model where light level and the number of strangled seedlings per pot were considered as fixed effects in a factorial design. Where the initial tests yielded F-scores < 0.05, Student's t-test or Tukey's HSD were used post-hoc, to perform pairwise comparisons within groups. The statistical analyses were carried out in JMP (JMP® pro 15.0.0, SAS Institute Inc.).

## FIELD EXPERIMENT

### The study site

The experiment was conducted in a 15-20-year-old, naturally regenerated *Pinus sylvestris* stand located in northern Sweden (64°14'N, 19°46'E, and 175 m above sea level). The soil is weakly podsolized sandy silt sediment, and the field layer consists mainly of lichens, with infrequent occurrences of *Calluna vulgaris* and *Vaccinium vitis-idaea*. The organic mor layer is 1-3 cm thick, and has a C-N ratio of  $37 \pm 1$  and pH of  $4.0 \pm 0.1$  (Hasselquist *et al.* 2016).



The trees were between three and five meters tall with stem diameters at breast height of  $7.4 \pm 2.6$  cm (mean  $\pm$  1 SD). The site is very N-poor and has an uneven stand density, including bald patches as well as patches where the trees grow closer together.

### Study design

We selected 23 circular plots with radius of two meters, using a tree as the center. All trees growing within this area were considered as part of the plot. Plots contained 5-11 trees ( $7.1 \pm 1.7$  mean  $\pm$  SE) and had a total basal area at breast height of  $3.2 \pm 1.2$  dm<sup>2</sup> (mean  $\pm$  SE). The plots were placed around trees growing in the denser patches of the site, so that they were naturally semi-discrete in the landscape. We had two reasons for doing this: First, the higher tree-density in these patches allowed us to assume that these trees were occupying the same soil volume and were more likely connected to the same mycorrhizal network. Second, the surrounding low-density areas should help reduce the influence from trees whose stems grew outside the plot boundary.

We designed four plot-level treatments using stem strangling to reduce the trees' belowground C-transport (detailed description in Henriksson et al., 2015). In Henriksson et al. (2015) canopy <sup>13</sup>C labelling and subsequent isotopic analysis of phloem carbohydrates showed that none of the <sup>13</sup>CO<sub>2</sub> absorbed after strangling was transported past the strangling point to the lower part of the stem. The experiment described in that publication was performed on similar trees to the current study, and in the same study area. Plots were considered to have two types of trees – the center tree, and neighbor trees –which could be either strangled or not strangled. In other words, we either strangled the stems of all trees, none of them, only the center tree, or all except the center tree (figure 2). The plots were divided into six blocks and then randomly assigned one of the four treatments. One of the blocks only had space for 3 plots, and thus one of the treatments (all trees strangled except one) was only replicated five times, but all the rest were replicated six times. In each plot, two trees were selected for needle sampling (3 weeks after <sup>15</sup>N-application), the central tree and one neighbor tree.

Mean basal area did not differ between plot treatments ( $p = 0.14$ , ANOVA), although the variance was unequal: the basal area of control plots varied more than the other treatments

( $p = 0.001$ , Levine's test). The number of trees per plot was not significantly different between treatments ( $p = 0.23$ , ANOVA).

### **Sampling and treatments**

All strangling treatments began on July 21<sup>st</sup>, 2015 (DOY 202). On August 10<sup>th</sup> (DOY 222), we applied 2.62 g of  $^{15}\text{N}$ -labeled  $\text{KNO}_3$  (0.39 g  $^{15}\text{N}$ ) dissolved in water, which could be detected in needle samples taken three weeks later. This N form was chosen for the high C requirement associated with its reduction and assimilation, which should lead to lower efficiency of N immobilization by free-living soil microbes than would be the case with N sources like ammonium or organic N. The application was equivalent to  $0.02 \text{ kg } ^{15}\text{N ha}^{-1}$  and the solution corresponded to  $2 \text{ liters m}^{-2}$ , which was evenly distributed from above and allowed to soak into the soil, over a circular area with a radius of 2.5 meters (plot radius + 0.5m), in order to treat a larger proportion of edge trees' root systems. The isotopic enrichment of N in the foliage of trees that received different treatments could then be compared to detect changes in uptake patterns among the treatments.

### **MODEL DESCRIPTION**

Our hypothesis is formulated in terms of a model describes the C-N exchange between plants and mycorrhizal fungi, both at the stand (or network) level and from the perspective of an individual plant. It was tested and evaluated based on the data from the pot experiment, which allowed better control and isolation, and more complete quantification of the  $^{15}\text{N}$  uptake by all plants than was possible in the field experiment. Strangling of a seedling predictably reduces its C provision below ground, to roots and the fungal network (Henriksson *et al.* 2015). Thus, we assume that the C supply to fungi is proportional to root biomass but that it is reduced by strangling according to a strangling factor estimated by the model.

### **Stand level C-N exchange**

The growth of mycorrhizal fungi is fueled by C supply by the plants ( $\dot{C}_s$ ), which we consider in relative terms (compared to mean of control plants) because its absolute value cannot be estimated and is not important for our conclusions. We assume that C supply to fungi from an individual plant ( $\dot{C}_{si}$ ) is proportional to its root mass ( $C_{ri}$ ) (Rouhier & Read 1998; Neumann

& Matzner 2013) and further reduced by strangling by a constant factor (see *supplementary information* for model parameterization).

We assume that fungal N uptake ( $\dot{N}_u$ ), is a saturating function of fungal growth (which is proportional to its biomass):

$$\dot{N}_u = \frac{N_a \dot{C}_s}{\dot{C}_s + \dot{C}_{sh}} \quad (1)$$

In eq. 2  $N_a$  = soil N availability (maximum potential N uptake) and  $\dot{C}_{sh}$  = half-saturation  $\dot{C}_s$ . N immobilization in fungal biomass is:

$$\dot{N}_f = \dot{C}_s \cdot I_f \quad (2)$$

The N immobilization factor,  $I_f = N:C_f \cdot e_f$ , where  $N:C_f$  = N:C ratio of fungal biomass and  $e_f$  = C use efficiency of fungal growth.

Combining eqs. 1 and 2, N export to plants ( $\dot{N}_p$ ), can be written as:

$$\dot{N}_p = \dot{N}_u - \dot{N}_f = \frac{N_a \cdot \dot{C}_s}{\dot{C}_s + \dot{C}_{sh}} - \dot{C}_s \cdot I_f \quad (3)$$

### Competition for N among individual plants

We assume that the fungi are attached to all plants in a pot and deliver N to each plant depending on its C supply relative to its competitors, which is postulated by eco-evolutionary theory (Wyatt *et al.* 2014) and indicated by experiments (Kiers *et al.* 2011; Fellbaum *et al.* 2014). This N competition effect was estimated in terms of N uptake of a plant  $i$  ( $\dot{N}_{pi}$ ) relative to mean N uptake of all plants in the pot ( $\overline{\dot{N}_p}$ ) as

$$\frac{\dot{N}_{pi}}{\overline{\dot{N}_p}} - 1 = d \cdot \left[ \left( \frac{\dot{C}_{si}}{\overline{\dot{C}_s}} \right)^z - 1 \right] \quad (4)$$

In eq. 4,  $\dot{C}_{si}$  = C supply from plant  $i$ ,  $\overline{\dot{C}_s}$  = mean C supply from all plants, and  $d$  = degree of fungal N export discrimination according to plant C supply.  $d$  is approximately equal to the marginal tree N gain per C supply for a tree, or the marginal C gain from each tree per N

export for a fungus. Theoretically,  $d=1$  maximizes the total C a fungus receives from all its tree partners, because a larger or smaller  $d$  would mean that the fungus could increase C supply by redistributing N supply among its tree partners. The parameter  $z$  allows for a potential non-linear effect of individual C supply on N uptake (for  $z \neq 1$ ), e.g. due to N limitation of fungal partners.

The modelled relationships between individual plant scale and network scale C and N exchange is illustrated in fig. 3. Detailed descriptions of parameter estimation and model testing are presented in the supplementary information for this article.

## RESULTS

We found that seedlings growing in shaded communities ( $78.8 \pm 4.8$  % reduction of PAR, mean  $\pm$  SD) for two months had 36 % smaller dry mass at the time of harvest than seedlings in sun pots ( $p < 0.0001$ , fig. 4a). Phloem strangling of individual seedlings did not affect their biomass, but it did cause a decrease in root/shoot ratio ( $p < 0.0001$ , fig. 4b).

Three weeks before harvest, 12 mg traceable nitrogen isotope  $^{15}\text{N}$  was applied to the soil surface of each pot. Shading led to 50% higher recovery of applied  $^{15}\text{N}$  in seedling biomass (1.07 mg  $^{15}\text{N}$  per seedling, compared to 0.71 mg  $^{15}\text{N}$  per seedling, fig. 4c;  $p < 0.0001$ ). Thus, a total of 54 % of applied  $^{15}\text{N}$  was accounted for in plant biomass in shaded pots, and only 36 % in sun pots. Further, strangled seedlings received significantly less  $^{15}\text{N}$  than non-strangled ones (fig. 4c). This supports the stoichiometric model of mycorrhizal C-N exchange, which predicts that decreased C export to fungi mobilizes soil N to the plant host and *vice versa*.

In shaded seedlings,  $^{15}\text{N}$  allocation to foliage was higher than for seedlings grown in full sun ( $p < 0.0001$ , fig. 4d). Strangling individual seedlings had the opposite effect on  $^{15}\text{N}$  allocation, compared to shading – more N remained in the mycorrhizal roots of the seedlings, which included both a fungal and a plant component ( $p < 0.032$ ).

In the field, we found that reducing belowground C flux, by strangling one tree per plot, improved total mobilization of applied  $^{15}\text{N}$  label to the trees (fig. 5a, b). In such plots, the strangled tree received less N than its neighbors (fig. S3). Strangling a greater subset of trees, thereby further impairing the capacity for plot-scale belowground C export, caused total  $^{15}\text{N}$  uptake to fall significantly, as was observed in plots where 82-100 % of tree basal area was strangled (fig. 5a, b).

Both shading and strangling reduce the belowground C flux that fuels the activity of mycorrhizal fungi. Based on the greenhouse experiment, we developed a model to test the connection between belowground C export and plant N acquisition at both the *individual tree-scale* (seedling) and at the *network-scale* (whole pot). The model explained 58% of the variation among individuals in the same pot, and 25% of the variation between pots in plant N uptake. In addition to the measured effects on root mass, the model indicated that C flux to mycorrhiza per root mass was significantly reduced by strangling (by 55%) but not by shading (Supplementary).

The model, supported by our measurements, shows that the greatest N-mobilization for plant use occurred at an intermediate level of belowground C export (fig. 6 a). Initially, N uptake and export to plants increases steeply with C supply to mycorrhizal fungi, but the rate of increase gradually declines as hyphae fill up the soil and N becomes limiting. At the same time N immobilization in fungal biomass increases linearly with C supply, which eventually results in declining net N export to plants. Thus, the model corroborates the hypothesis that maximum *network-scale* N mobilization occurs at an intermediate level of *network-scale* C export. At the individual tree-scale, the marginal increase in the share of N that the plant receives per share C supplied is approximately equal to one (0.95), as theoretically predicted (fig. 6b). This drives a competition for N among trees where each tree at first gains N by fueling fungal growth, but later the whole community suffers from N immobilization in the shared fungal network.

## DISCUSSION

We show that belowground C allocation to can fuel N immobilization, reducing the amount of N to be distributed among the trees. But we also found that individual trees received nutritional benefits in proportion to their carbon contribution to the fungal network in accordance with our hypothesis (fig. 1, alternative 3). This apparent incongruity can be explained by invoking the concept of the *tragedy of the commons*, as described by Hardin in 1968 (Hardin 1968).

Our estimates of plant C export to EMF are constrained by root measurements and a strangling effect (previously shown to predictably reduce below ground C export (Henriksson et al 2015)). The only free parameter affecting the modelled C export was the magnitude of the strangling effect per root biomass. As an additional test, we used respiration measurements to make independent estimates of the C export to EMF at the pot (community) level, which were well correlated the model results, and suggesting that the model explains 39% of the variation in EMF respiration (Supplementary information). The underlying details of the strangling effect on C export are not relevant for our conclusions but may include reduced EMF colonization of roots (Björkman 1944), and reduced growth of extraradical mycelia extending from strangled roots. Either way, the strangled plants cannot have been completely ejected from the EMF network, or their N uptake would not fall on the same line as the non-strangled plants in figure 6b.

The most plausible alternative scenarios and their implications are displayed in fig. 1: (1) direct N uptake without EMF and (2) N uptake via EMF but without a common network. In alternative 1, plant N uptake would increase with below ground C allocation to roots and would tend to saturate, but never decline, at higher C allocation. Competition would lead to slightly less saturation at the individual than at the community level (Franklin et al 2012). Alternative 2 would lead to initially increasing but eventually saturating plant N uptake with C export, due to linearly increasing fungal growth and N immobilization with C export. The response would be similar at both the individual level and the community level as there is no strong inter-plant competition for EMF-derived N. Only if plants take up and compete for N via a common EMF network (Alternative 3 in fig. 1) is it possible to obtain the contrasting results shown in fig. 6, i.e. a linear increase in plant N uptake with C export at the individual level (scaling exponent = 1.038, Supplementary) and a hump shaped relationship at the

community level. The linearity of the individual response was tested, resulting in a. We conclude that the most reasonable interpretation of this data is the presence of common EMF network (Nara 2006; Beiler *et al.* 2010) in which multiple fungi connect the host plants to each other (Franklin *et al.* 2014).

A stable evolutionary strategy for a multi-partner trade network is achieved when individuals allocate resources among symbionts of the other species in proportion to the relative benefits they receive from each partner. This is called proportional discrimination and has been applied to modelled mycorrhizal networks (Wyatt *et al.* 2014). The fact that our model (where plant N uptake was measured and relative C contribution to fungi was modelled) resulted in a linear proportionality (with slope 0.94, not significantly different from 1) between relative C investment and N uptake strongly indicates the presence of a common mycorrhizal network.

In our experiment, seedlings were potted in soil containing inoculum of the EMF *Suillus variegatus*. This was done to ensure a minimum degree of comparability among the plant-fungal systems established in the pots. As mentioned in the Methods section, seedlings were 2 years old at the time of planting, and any EMF species already present on the root systems from their time at the nursery were not excluded. We consider this to be a strength of the current study, as the presence of multiple fungal species improves the comparability of the results to those of the field study. Further, mycorrhizal market theory (Franklin *et al.* 2014; Wyatt *et al.* 2014), upon which our hypotheses were based, does not rely on species differences or a particular species being present, it is the competition between multiple individuals (regardless of species) that drives the dynamics.

Rising atmospheric CO<sub>2</sub> could significantly increase mycorrhizal fungal biomass (Treseder 2004), which could drive progressive N limitation in forests via mycorrhizal immobilization (Alberton *et al.* 2007; Alberton & Kuyper 2009; Steidinger *et al.* 2019). Our results support this notion. In fact, the field-studied Scots pines were observed to export higher-than-optimal quantities of C under untreated conditions (fig. 5). Therefore, any increase in the C supply to EMF should further exacerbate the network-scale N limitation in the studied forest stand (fig. 3, 6a). However, this may not be the case in locations where soil N availability is

greater, or in situations where increased C supply allows EMF to have access to more energy-demanding N sources, such as complex organic substrates.

Although a global model meta-analysis of elevated CO<sub>2</sub> experiments (Terrer *et al.* 2019) concluded that rising atmospheric CO<sub>2</sub> would continue to stimulate plant growth in boreal forests in general, empirical evidence for N-poor boreal forests is scant. Of the two experiments from forests similar to ours included in the meta-analysis, one showed a small negative CO<sub>2</sub> effect (Sigurdsson *et al.* 2013). In support of our prediction, Alberton *et al.* (2005) observed that, in laboratory conditions, the growth-enhancing effect of elevated CO<sub>2</sub> was greater in the fungal component of an ectomycorrhizal symbiosis than it was in the plant. They concluded that this should eventually cause increased plant-fungus competition for N, and suggested this as a mechanism for future progressive N limitation, in line with our model predictions. This further suggests that the risk of losing forests dominated by ectomycorrhizal tree species due to climate change (Steidinger *et al.* 2019) may be unfounded, as that study did not account for increasing N limitation (relative to C) caused by rising CO<sub>2</sub>, which should reinforce the stability of the ectomycorrhizal symbiosis (Franklin *et al.* 2014).

That the *tragedy of the commons* mechanism has gone almost unnoticed by scientists until now may be due to many mycorrhizal C-N trade experiments having used setups containing a single plant, paired with a fungal partner (Colpaert *et al.* 1996; Corrêa *et al.* 2010). Such a design cannot capture network-scale drivers of EMF-plant interactions. Arbuscular mycorrhiza connected to multiple host plants can preferentially supply nutrients to the plants presenting greater C-sources for the fungi (Fellbaum *et al.* 2014; Weremijewicz & Janos 2013; Weremijewicz *et al.* 2016). However, the current study is the first to show the link between plant-plant competition within EMF networks and the resulting N immobilization as measured at the system level.

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